

## Full curriculum

### Professional Master's Degree – Specialty: Biotechnology and Applied Microbiology

**Semester: S1**

**Fundamental Teaching Unit: UEF1(O)**

**Course Title: Rhizobial Symbioses**

**Credits: 6**

**Coefficient: 3**

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#### **Course Objectives**

By the end of this course, students will be able to:

- **Study symbioses** involving **nitrogen-fixing bacteria and leguminous plants**.
- **Identify symbiosis types** and analyze **morphological and physiological characteristics** of these interactions.
- **Apply theoretical knowledge** through **practical lab work (TP)** to reinforce the study of these symbioses.

#### **Recommended Prerequisites**

- Knowledge of **different types of actinorhizal associations** and their role in **ecosystem stability and soil protection**.
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#### **Course Content**

Lectures

1. General Overview
  2. Overview of Nitrogen-Fixing Bacteria
  3. Diversity and Taxonomy of Nitrogen-Fixing Bacteria
  4. Plant Partners
  5. Bacterial Partners
  6. Establishment of Symbiosis
  7. Molecular Dialogues Between Symbiotic Partners
    - Mechanisms of symbiosis establishment
    - Nodule formation and regulation
  8. Temperate Symbioses
  9. Tropical Symbioses
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#### **Practical Work (Lab Sessions)**

- I. Isolation of Bacteria from the Rhizosphere of Legumes
- II. Macroscopic Study
- III. Microscopic Study
- IV. Characterization of Cultural Traits
- V. Inoculation of Germinated Seeds
- VI. Infectivity Tests

Semester: S1

Fundamental Teaching Unit: UEF1(O) – Symbiotic Microorganisms

Course Title: Actinorhizal Symbioses

Credits: 6

Coefficient: 3

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### Course Objectives

- Understanding different types of actinorhizal associations and their role in ecosystem stability and soil protection.

### Recommended Prerequisites

- General microbiology (L2 level)
  - Environmental microbiology
  - Microorganisms of interest (L3 level)
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### Course Content

I. Symbiotic Partners in Actinorhizal Symbiosis

II. Bacterial Partner: Actinomycetes of the Genus *Frankia*

III. Isolation and In Situ Characterization

IV. Morphology and Cultural Characteristics

V. Diversity and Taxonomy of *Frankia*

VI. Saprophytic Lifestyle of *Frankia* in Soil

VII. Host Plants

VIII. Infection and Nodulation in *Frankia*

IX. Two Modes of Infection in *Frankia*

X. Development and Structure of Nodular Lobes

XI. Molecular Signals Involved in Nodulation

XII. Genetic Control of Actinorhizal Symbiosis

XIII. Biotechnological Importance of Actinorhizal Symbiosis

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## Evaluation Method

- Continuous assessment
  - Final exam
  - Personal research and oral presentation
  - Evaluation of practical work (TPs)
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## Independent Study and Assignments

- Research on key concepts and mechanisms of actinorhizal symbiosis, assigned after each lecture.
- Preparation of technical sheets for various practical sessions, including:
  - Purification of *Frankia* strains
  - Preparation of inoculum
  - Inoculation tests
  - These sheets should be based on bibliographic research and will be corrected and evaluated.
- Students will submit their lab reports as mini-theses.

## Semester: S1

### Fundamental Teaching Unit: UEF1(O) – Symbiotic Microorganisms

**Course Title: Mycorrhizal Symbioses**

**Credits: 6**

**Coefficient: 3**

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## Course Objectives

- Understanding the **mechanisms of establishment, regulation, and functioning** of mycorrhizal associations.
- Recognizing the **importance of mycorrhizae** in plant development and **ecosystem stability**.

## Recommended Prerequisites

- Basic knowledge of **general microbiology (L2 level), environmental microbiology, fungi, and mycorrhizae**.
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## Course Content

Lectures

- I. Major Types of Mycorrhization and Their Organization
  - II. Classification of Mycorrhizae
  - III. Endomycorrhizae
  - IV. Life Cycles of Mycorrhizal Fungi
  - V. Infection of the Host Plant
  - VI. Arbuscules and Vesicles
  - VII. Cellular and Molecular Modifications During Mycorrhizal Development
  - VIII. Pseudonodules
  - IX. Ectomycorrhizae
  - X. Mycorrhizae and Mushroom Fruiting
  - XI. Plant-Fungus Interactions
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### Practical Work (Lab Sessions)

- I. Search and Isolation of Mycorrhizae from Different Biotopes
  - II. Detection of Endomycorrhizal Structures
  - III. Identification of Endomycorrhizal Structures
  - IV. Microscopic Observation of Endomycorrhizal Structures
  - V. Evaluation of Mycorrhization Rate and Frequency
  - VI. Spore Extraction
  - VII. Description of Mycorrhizal Structures (Arbuscules, Vesicles)
  - VIII. Spore Counting and Identification
  - IX. Assessment of Mycorrhization Frequency and Rate
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### Independent Work and Research Topics

- Study on the feasibility of producing and commercializing fungi as biofertilizers.
  - Development of a mini-project on the production and commercialization of edible mushrooms.
  - Market potential analysis of mycorrhiza-based biofertilizers.
  - Economic impact study of biofertilizer commercialization at a regional scale (in collaboration with agricultural stakeholders).
  - Final project presentation before the academic committee, evaluated at the end of the semester.
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### Evaluation Method

- Continuous assessment
- Final exam
- Independent research, bibliographic study, and oral presentation

**Semester: S2**

**Fundamental Teaching Unit: UEF1**

**Course Title: Methods for Evaluating the Efficiency of Microbial Symbioses**

**Credits: 3**

**Coefficient: 6**

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### **Course Objectives**

- Acquire **practical techniques for estimating symbiotic nitrogen fixation**, enabling the assessment of **soil fertility** and its **potential for agricultural valorization**.

### **Recommended Prerequisites**

- Knowledge of **physiology and biochemistry**.
    - Understanding of **plant-associated microorganisms** (acquired in L3).
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### **Course Content**

- I. Overview of Microbial Symbioses
  - II. Mechanisms of Nitrogen Fixation (*Functioning of Nitrogenase*)
  - III. Different Estimation Methods
  - IV. Measurement of Acetylene Reduction Activity (ARA)
  - V. Measurement of Fresh and Dry Weight
  - VI. Total Nitrogen Quantification
  - VII. Protein Quantification
  - VIII. Estimation of Ureides
  - IX. <sup>15</sup>N Isotope Labeling Technique
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### **Independent Work and Research Topics**

- Research on nitrogen fixation mechanisms and related concepts.
  - Preparation of technical sheets for various practical sessions, including:
    - Inoculation in hydroponic and solid media
    - Indirect evaluation methods (dry weight estimation with statistical analysis)
    - Protein quantification methods
  - Bibliographic research for establishing experimental protocols and methodological approaches.
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### **Evaluation Method**

- Final exam
  - Continuous assessment
  - Oral presentations
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### **References**

- Books, academic papers, and online resources (to be specified by the faculty team).

**Semester: S2**

**Fundamental Teaching Unit: UEF1**

**Course Title: Endophytes – Diversity and Role in Microbial Symbioses**

**Credits: 3**

**Coefficient: 6**

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## **Course Objectives**

- Discover and apply **microorganisms living within plant tissues** that play a crucial role in **enhancing plant growth**.

## **Recommended Prerequisites**

- Basic knowledge in **microbiology and plant-microbe interactions**.
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## **Course Content**

- I. Introduction and Historical Background
  - II. Different Groups of Endophytes (Bacteria and Fungi)
  - III. Mechanisms for Plant Growth Promotion (PGP)
  - IV. Isolation and Study Techniques
  - V. Characterization, Taxonomy, and the Role of Molecular Tools in Endophyte Identification
  - VI. Endophyte Colonization Mechanisms
  - VII. Associated Plant Partners – Selection Mechanisms
  - VIII. Fundamental and Applied Importance of Endophytes
  - IX. Association with Legumes
  - X. Association with Grasses (Case Study: Rice)
  - XI. Influence of Environmental Factors on Endophyte Growth and Efficiency
    - a. Biotic Factors
    - b. Abiotic Factors
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## **Independent Work and Research Topics**

- Research on Endophyte Colonization Mechanisms and techniques for in planta visualization of microorganisms.
  - Preparation of technical sheets for practical sessions, including:
    - Isolation and purification of endophytic strains
    - Characterization and inoculation testing
  - Oral presentations on molecular biology techniques used for endophyte identification.
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## **Evaluation Method**

- Continuous assessment
- Final exam
- Oral presentations and research reports

**Semester: S2**

**Fundamental Teaching Unit: UEF2**

**Course Title: Microorganisms and the Environment**

**Credits: 3**

**Coefficient: 6**

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## **Course Objectives**

- Provide students with a **comprehensive understanding of microorganism-environment interactions.**
- Explore the **role of microbial communities** in maintaining ecosystem balance and their **spatiotemporal evolution.**

## **Recommended Prerequisites**

- Basic knowledge of **microbial physiology, biochemistry, and general microbiology.**
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## **Course Content**

### **I. Introduction and Historical Overview**

### **II. Microbial Ecology Overview**

### **III. Economic and Environmental Importance of Microorganisms**

### **IV. Spatiotemporal Evolution of Microorganisms in an Ecosystem**

### **V. Habitat Study and Distribution of Bacterial Species**

- Cyanobacteria and Anoxygenic Photosynthetic Bacteria

### **VI. Microbial Diversity in the Environment**

### **VII. Identification Methods**

- Fluorescent Antibody Techniques
- Fluorescent **In Situ** Hybridization (FISH)
- Microautoradiography

### **VIII. Microbial Activities in the Environment**

### **IX. Quantification Methods for Microbial Activities**

### **X. Microbial Population Density and Activity**

### **XI. Concepts of Microbial Niches, Microenvironments, and Micro-Ecosystems**

## **XII. Microorganisms and Fundamental Biogeochemical Transformations**

## **XIII. Microbial Communities**

- **Primary Producers**
- **Consumers**
- **Decomposers**

## **XIV. Biofilms, Microbial Mats, and Biomass: Economic and Environmental Importance**

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### **Practical Work (Lab Sessions)**

- **Isolation of Microorganisms** from Different Environments
  - **Purification of Isolates**
  - **Study of Microbial Diversity** in Isolated Populations
  - **Partial Characterization of Microbial Strains**
  - **Preservation of Pure Strains**
  - **Study of Microbial Interactions**
  - **Antibiosis Studies**
  - **Analysis of Proteolytic Activities**
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### **Independent Work and Field Activities**

- **Field Visits:**
    - Wastewater treatment plants
    - Waste valorization facilities
  - **Project Presentation:**
    - Proposal for solving an environmental issue
    - Compost preparation
    - Waste valorization techniques
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### **Evaluation Method**

- **Final Exam**
- **Continuous Assessment**
- **Oral Presentations**

**Semester: S3**

**Fundamental Teaching Unit: UEF2 – Biotechnological Applications**

**Course Title: Microbial Inocula – Production, Quality Control, and Inoculation Technology**

**Credits:** 6  
**Coefficient:** 3

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## Course Objectives

By the end of this course, students will:

- **Master the techniques for producing and applying microbial inocula** (biofertilizers) in controlled mycorrhization.
- **Ensure quality control and monitor the sustainability of biofertilizers** while adhering to legal regulations and commercialization standards.

## Recommended Prerequisites

- Knowledge of **symbiotic microorganisms and their importance** (L3 level).
  - Understanding of **mycorrhizal fungi** (*Endomycorrhizae* and *Ectomycorrhizae*).
  - Familiarity with **legume-nodulating bacteria** (L3 level).
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## Course Content

### I. Introduction to Inoculation

- Definition of inoculation
- When and why inoculation is necessary
- Objectives of inoculation

### II. Types of Inocula

### III. Selection and Conditioning of Inoculum Carriers

### IV. Inoculum Preparation

- Characteristics of a **high-quality inoculum**
- **Microbial growth** and its study methods
- **Laboratory-scale production**
- **Industrial-scale production**
- **Bioreactors:** Description and operating modes
- Types of bioreactors used in microbial biomass production
- **Preservation of microbial inocula**

### V. Microbial Inoculum Quality Control

- **Microbiological control**
- **Genetic control**
- **Quality standards and norms**

## **VI. Legislation, Commercialization, and Management of Microbial Inocula**

## **VII. Inoculation Techniques**

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### **Practical Work (Lab Sessions)**

- I. Isolation of Rhizosphere Microorganisms (Rhizobia, Mycorrhizae, PGPR, etc.)**
  - II. Identification and Selection of Microbial Inocula**
  - III. Selection of Carriers (peat, waste, compost, etc.)**
  - IV. Efficacy Testing at Lab and Nursery Scale**
  - V. Quality Control and Long-Term Monitoring of Inocula**
  - VI. Demonstration and Field Testing with Farmers**
  - VII. Inoculum Formulation and Packaging**
  - VIII. Preservation Methods**
  - IX. Approval and Certification (Homologation)**
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### **Independent Work and Research Topics**

- Study of **commercialized microbial inocula**.
  - Research on **different carriers used for inoculum preservation**.
  - Selection of **bioreactors used in microbial inocula production**.
  - Effect of **inoculation on plant productivity**.
  - Impact of **inoculation on soil bioremediation**.
  - Influence of **inoculation on soil microbial flora**.
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### **Evaluation Method**

- **Final exam (EMD)**
- **Continuous assessment**
- **Technical reports**
- **Oral presentations**